

## Abstract View

## HIGH SPATIAL AND TEMPORAL RESOLUTION ESTIMATES OF CALCIUM DYNAMICS IN DENDRITIC SPINES USING MCELL SIMULATIONS.

[K.M. Franks<sup>1,2\\*</sup>](#); [T.M. Bartol<sup>1</sup>](#); [M.M. Poo<sup>2</sup>](#); [T.J. Sejnowski<sup>1,3</sup>](#)

1. CNL, Salk Inst, La Jolla, CA, USA
2. Biology, UCSD, La Jolla, CA, USA
3. HHMI, La Jolla, CA, USA

Calcium plays a critical signaling role in many biological systems. For example, in the postsynaptic neuron, increases in  $\text{Ca}^{2+}$  concentration are believed to be essential for the induction of long-term potentiation and long-term depression, changes in synaptic efficacy thought to be the cellular mechanisms underlying learning and memory. Measurements of intracellular  $\text{Ca}^{2+}$  have typically relied on the use of fluorescent imaging where the sensors used buffer the  $\text{Ca}^{2+}$ , obscuring concentration measurements, and have limited spatial and temporal resolution. In postsynaptic spines, the spatial localization of  $\text{Ca}^{2+}$ -dependent effectors such as calmodulin, CaMKII and calcineurin make the measurement of  $\text{Ca}^{2+}$  concentration in the entire spine less than ideal. Using MCell, a Monte Carlo simulator, to monitor both the influx of  $\text{Ca}^{2+}$  into a spine and dendrite and its reactions with intracellular molecules, we have reproduced fluorescent transients measured in neocortical and hippocampal neurons. This has allowed determination of the unperturbed  $\text{Ca}^{2+}$  dynamics and suggests kinetics and concentrations of endogenous  $\text{Ca}^{2+}$  binding proteins. We have measured  $\text{Ca}^{2+}$  concentrations in microdomains throughout the spine showing highly heterogenous concentrations of free intracellular  $\text{Ca}^{2+}$ . Furthermore, we show the spatial and temporal activation of different  $\text{Ca}^{2+}$ -dependent effectors resulting from asynchronous pairing of pre- and postsynaptic action potentials. To complement experimental studies, we show how computer simulations can offer an alternate and complementary method of assaying intracellular  $\text{Ca}^{2+}$  dynamics and reactions.

Supported by: NSF, HHMI, NIH & HFSP.



Site Design and Programming © ScholarOne, Inc., 2000. All Rights Reserved. Patent Pending.